# **Catalytic Peroxidation as a Feasible Approach to O-Phthalaldehyde (OPA) Oxidation in Wastewater**

ORAL Ph.D. Student: N Journal: JECE

*L. Doumic1,2, M. Antunez<sup>1</sup> , A.M. Ferro Orozco<sup>1</sup> , M.A. Ayude1,2. (1) INTEMA, CONICET, UNMdP, Av. Colón 10850, Mar del Plata, Argentina, luciladoumic@fi.mdp.edu.ar. (2) DIQyA, Universidad Nacional de Mar del Plata (UNMdP), Argentina.*



Improper discharge of disinfectants to sewarage and wastewater treatment plants can cause acute intoxication and chronic lowdose exposure, which can promote the development of antibiotic resistance. Ortho-phthalaldehyde (OPA) is a commonly used highlevel disinfectant for heat-sensitive, semi-critical medical devices in hospitals and clinical establishments, which is frequently discharged to the sewer network without a proper treatment. The aim of this study was to evaluate for the first time the feasibility of treating OPA with catalytic peroxidation processes. The homogeneous Fenton oxidation, along with the heterogeneous Fenton-type process using a catalytic bed of Prussian Blue nanoparticles supported on alumina spheres (PBNP/ $\gamma$ -Al<sub>2</sub>O<sub>3</sub>) were explored. Results demonstrated that, under appropriate operating conditions, a 180 min treatment could be effective to achieve a free OPA and non-toxic solution, with a low Dissolved Organic Carbon content, no residual  $H_2O_2$ , and no sludge.

### **Introduction**

There is increasing concern regarding water quality due to emerging contaminants entering ecosystems through untreated effluent discharge. The extended especially in healthcare establishments to combat pathogens, poses risks of<br>environmental contamination and disease contamination dissemination. Improper disinfectant application can lead to sub-lethal concentrations, microbial resistance and decreased effectiveness of antimicrobial agents. To address the limitations of traditional biological treatments in the presence of toxins, innovative solutions are needed for the safe disposal of disinfection byproducts.

Among Advanced Oxidation Treatments, the Fenton process stands out for its simplicity, gentle or moderate operating conditions, cost-effectiveness, availability, and low toxicity of reagents [1]. However, its implementation often faces challenges such as potential formation of insoluble complexes between iron ions and organic matter, and the generation of sludge, which limit the treatment success [2, 3]. Alternatively, the use of heterogeneous catalysts can mitigate these issues by reducing sludge formation and allowing easier recovery and reuse. This study explores, for the first time**,** the feasibility of using catalytic peroxidation processes to oxidize commercial solutions of o-phthalaldehyde (OPA), a commonly used high-level disinfectant for heatsensitive, semi-critical medical devices in hospitals and clinical establishments .

## **Material and Methods**

The Homogeneous Fenton (HF) experiments were conducted in a thermostated agitated batch reactor. The reactor was filled with 150 mL of OPA solution at a known concentration and  $pH = 3$ . Once the desired temperature was reached, the reaction was initiated by adding ferrous sulfate and  $H_2O_2$ .

The Heterogeneous Fenton-type (HFT) oxidation was investigated using a liquid batch-recycle reactor. The reactor consisted of an 11 mm ID glass column containing 5 g of randomly packed "insoluble"  $PBNP\bar{Y}_1$ -Al<sub>2</sub>O<sub>3</sub> catalyst, prepared as described in [4]. To start each test, 200 mL of OPA solution with a known concentration and  $pH = 3$  were introduced into a reservoir.  $H_2O_2$  was added and allowed to mix. The initial sample was collected, and the liquid flow through the reactor, set at 2.2 L/h, was initiated using a peristaltic pump. Temperature was maintained at 70ºC by submerging the system in a thermostatic bath.

In both, HFT and HF processes, liquid samples were periodically taken out from the reservoir or the reactor and analyzed at once in terms of pH, UV–Vis spectrum, OPA,  $H_2O_2$ , total dissolved iron (TDI) and Dissolved Organic Carbon (DOC) concentrations.

Analytical techniques:  $H_2O_2$  concentration was determined by a Glycemia enzymatic test (Wiener Lab.). OPA concentration was determined by gas chromatography in a Hewlett-Packard 5890 series II, equipped with a FID detector and a capillary column. TDI concentration was assessed by measuring the total iron content of filtered samples with the ophenantroline method [5]. DOC was measured in a Shimadzu TOC-VCPN analyzer. Shimadzu UV-1800 spectrophotometer was used to obtain the UV–Vis spectrums between 190 and 800 nm. The carboxylic acids formed were quantified by ion-exclusion in an HPLC (Shimadzu LC-20AT) equipped with a Rezex ROA-Organic Acid H+  $(8 \%)$  300  $\times$  7.8 mm (Phenomenex) column. Toxicity was assessed by measuring the inhibition of respiratory activity  $(I_{\text{SOLR}})$ of activated sludge system (ISO 8192 method).

### **Results and Discussion**

Different homogeneous alternatives for treating OPA 1000 mg  $L^{-1}$  (DOC = 700 mg<sub>DOC</sub>  $L^{-1}$ ) were evaluated. The order of addition of Fenton reagents was crucial for the feasibility of HF treatment, with the oxidant needing to be added last. Using 80 mg L-1 of Fe+2 and the stoichiometric dose of  $H_2O_2$  necessary to mineralize initial OPA, yielded excellent results (Figure 1).



**Figure 1.** Temporal profiles of (a) normalized  $H_2O_2$ concentration and (b) DOC conversion along the temperature-controlled process at T=(■) 30°C, (▲) 50°C and ( $\bullet$ ) 70°C for 180 min. Inset: Profiles during the storage stage at room temperature, without stirring and in darkness. Conditions:  $[OPA]_0 = 1000$  mg L<sup>-1</sup>, pH<sub>0</sub> = 3,  $[Fe^{+2}]_0 = 80$  mg L<sup>-1</sup>, [H<sub>2</sub>O<sub>2</sub>]<sub>0</sub> = 135 mmol L<sup>-1</sup>

Complete  $H_2O_2$  consumption, 100% OPA degradation, 77-85% mineralization and no toxicity  $(I_{\text{SOUR}} \le 0\%)$  were achieved. Sludge formation was observed accompanied by a low oxidant accompanied by a low oxidant consumption rate. The formation of carboxylic acids as intermediates, promoted a gradual increase in TDI concentration, favoring oxidant consumption and DOC conversion. The time required to attain these results depended on the operating temperature during the first 180 min (30°C, 50ºC or 70°C). A storage stage (without stirring, dark and room temperature) was needed for reactions conducted at 30 and 50 ºC, whereas a 180 min treament was enough at 70ºC. These results indicate the potential for further investigation into HFT oxidation. The catalyst based in supported Prussian blue nanoparticles (PBNP/γ- $Al_2O_3$ ) showed to be active in the HFT oxidation of OPA solutions (Figure 2). A solution of 1000 mg  $L^{-1}$ 

could not be effectively treated with 5 g of catalyst, resulting in a solution with toxicity ( $I_{\text{SOLR}} = 76\%$ ), despite 57% of mineralization was attained. In contrast, 95% of mineralization and a non-toxic solution ( $I_{SOUR} = 0\%$ ) was achieved when treating a solution of 100 mg  $L^{-1}$  in 180 min. These findings demonstrate that a non-toxic solution can be achieved under appropriate operating conditions.



**Figure 2.** Evolution of normalized H<sub>2</sub>O<sub>2</sub> concentration and DOC conversion in the HFT oxidation of solution with [OPA]<sub>0</sub> (•) 1000 mg L<sup>-1</sup> and ( $\star$ ) 100 mg L<sup>-1</sup>. [H<sub>2</sub>O<sub>2</sub>]<sub>0</sub> = 14 mmol L<sup>-1</sup> .

## **Conclusions**

This study demonstrated the feasibility of treating OPA solutions with highly oxidizing species generated in AOPs. Further work is being conducted to identify suitable operating conditions to effectively treat hospital disinfection wastewater to ensure its safe disposal into the sewer network.

#### *Acknowledgments*

The authors gratefully acknowledge financial support from the National University of Mar del Plata, the CONICET (PIP 1220200102822CO, PIBAA-2022–2023-28720210100663CO) and the ANPCyT (PICT-2020-SERIEA-03170).

#### *References*

- **[1]** A. Babuponnusami, y K. Muthukumar, *J. Environ. Chem. Eng.*, 2 (1) (2014) 557.
- **[2]** L. Doumic, P. Soares, M. Ayude, M. Cassanello, R. Boaventura,V. Vilar, Chem. Eng. J., 277 (2015) 86.
- **[3]** C. Benatti, A. Saraiva da Costa, C. Granhen Tavares, J. Hazard. Mater., 163 (2009) 1246.
- **[4]** L. Doumic, G. Salierno, C. Ramos, P. Haure, M. Cassanello, M. Ayude. RSC Advances, 6 (2016) 46625.
- **[5]** Colorimetric Determination of Traces of Metals, E.B. Sandell (3rd edition), New York, Interscience Publishers Inc, 1959.